

# UCL Institute of Child Health

Dr Bertrand Vernay  
Light Microscopy Facility Manager

42224

Tel Mobile: 07582249337

[Email: b.vernay@ucl.ac.uk](mailto:b.vernay@ucl.ac.uk)

Revised December 2014

# Table of contents

page 3.....Ownership

page 3..... Access Rules

page 3.....Olympus Customer Support Contact

page 4 .....General Specifications

Microscopy Techniques Available

Objectives

Filter Cubes

Prof. Jane Sowden, Developmental Biology Unit (Purchased in 2011)

No access without prior training by the Light Microscopy Facility Staff  
Free of hourly charge for Sowden and Ferretti groups, £1 hourly charge for  
all other users towards the cost of the consumables is expected  
Prof. Sowden team has priority over other users.  
Users must always record their activity in the Log book  
Problem(s) with the microscope should be reported as soon as they are  
noticed

## General specifications

Microscopy techniques available

### Objectives

Olympus UPlanFLN 10x Ph1 NA 0.3 WD 10.0 mm

Olympus LUCPlanFLN 20x Ph1 NA 0.45 WD 6.6-7.8 with correction collar

Olympus LUCPlanFLN 40x Ph2 NA 0.6 WD 3.0-4.2 with correction collar

Zeiss objectives can also be used

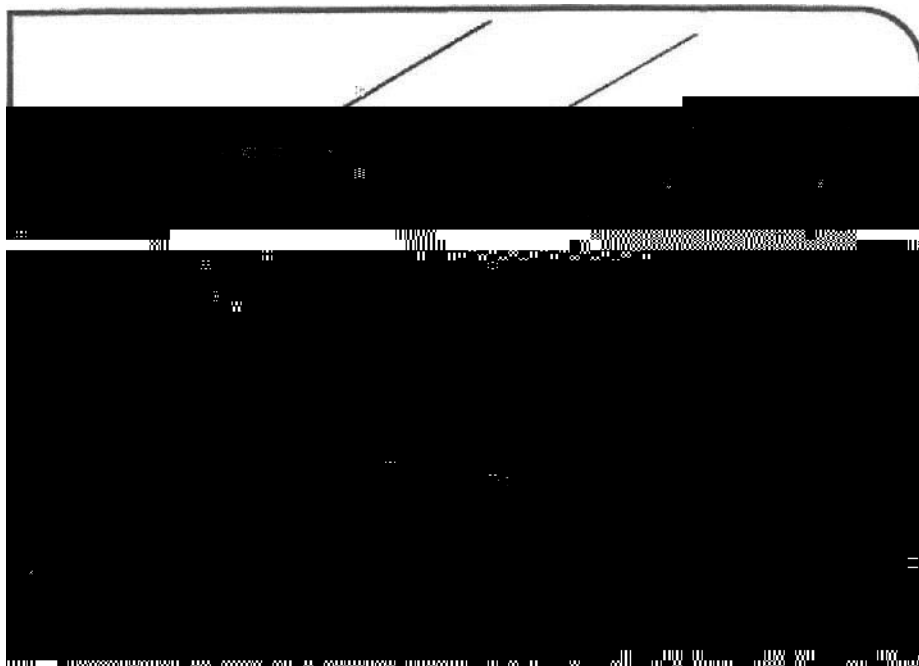
### Filter cubes (see p11-15)

Position	Filter set name	Exciter	Beamsplitter	Emission
1	DAPI	350/50x	400LP	



## Halogen lamp operation: Turning on the lamp

1. Make sure the light intensity control knob (5) is in the MIN (minimum intensity) position on the microscope frame.
2. Make sure the light intensity control knob (1) is in the MIN (minimum intensity) position on the TH4 module.
3. Set the main switch (2) to "I" (ON) on the TH4 module.
4. On the microscope front, press the transmitted light ON-OFF button (6) so that the button is illuminated.
5. Adjust the brightness with the light intensity control knob (5).
6. To turn OFF, set the transmitted light ON -OFF button (6) to OFF



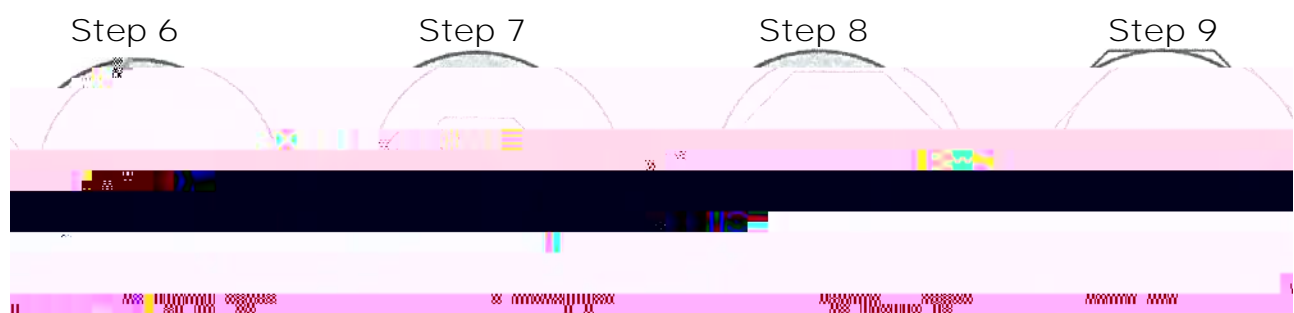
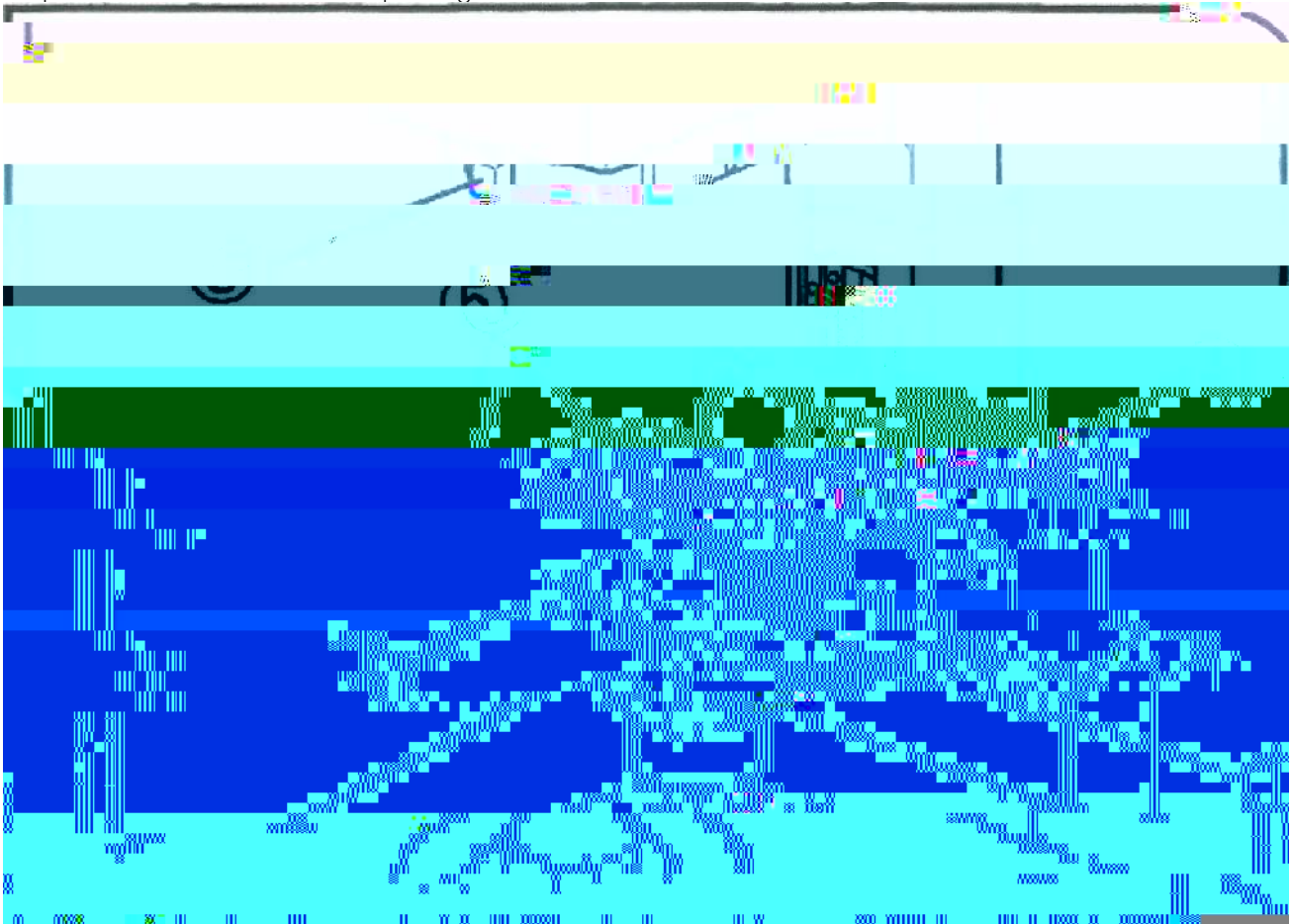
## Halogen lamp operation:

### Turning off the lamp

1. Set the light intensity control knob (5) to the MIN (minimum intensity) position on the microscope frame.
2. Set the light intensity control knob (1) to the MIN (minimum intensity) position on the TH4 module.
3. Set the main switch (2) to "0" (OFF) on the TH4 module.

## Kohler illumination:

1. Rotate the turret (1) to the "BF" position. (Any of positions 3,4 or 5, position 1=Ph1, 2 = Ph2)
2. Slide the aperture iris diaphragm lever to fully open the diaphragm.
3. Slide the field iris diaphragm lever to the fully open position.
4. Engage the 10x objective and bring the specimen into focus.
5. Using the field iris diaphragm lever, completely close the field iris diaphragm.
6. Rotate the condenser height adjustment knob (4) to bring the field iris diaphragm image into focus.
7. Center the field iris diaphragm using the condenser centering knobs (5).
8. Open the field iris diaphragm until its image reach the limits of the field of view, adjust the centering if necessary.
9. Open the field iris diaphragm until not visible.











Position #1

49000 350 0 0 (en/>B5 (en0 (enxETBT1.0364 0408.0364 01 9.e)-802[(P)0 1 10ETBT

---

Position #2  
 41017 EndowGFP/EGFP Bandpass  
 Emitter ET470/40x  
 Beamsplitter 495LP  
 Emitter ET 525/50m

Wavel ength (nm)

H0470/40x  
 H0525/50m

Q4951p

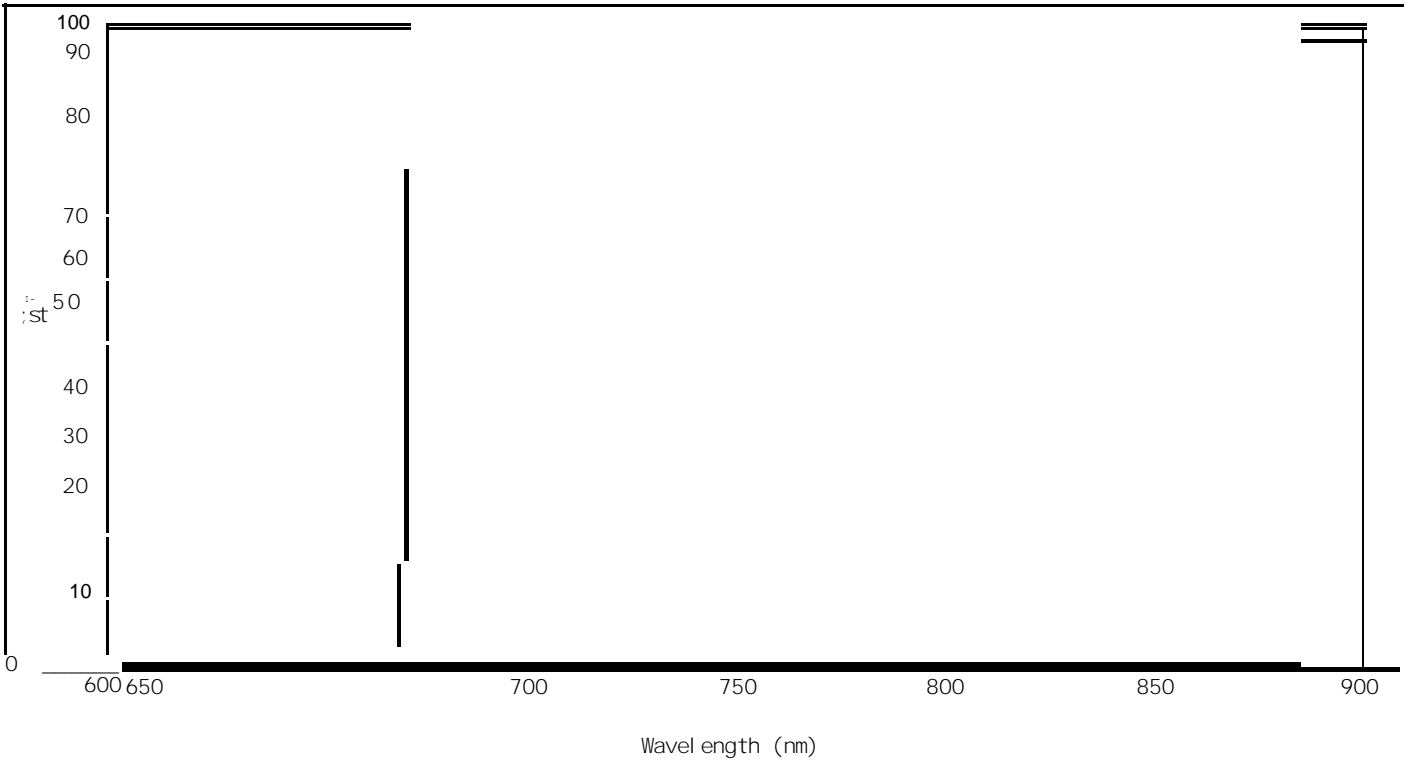
Fluorochrome	EX	EM	Use
Acridine Orange + DNA	500	526	Alternative
Alexa Fluor 488T"	498	520	Alternative
Azami Green	492	505	Alternative
BODIPY FL/pH7.2	505	512	Alternative
Calcein	494	517	Alternative
Calcium GreenTM - 1	506	531	Alternative
Cy2TM	489	506	Alternative
DiO	484	502	Alternative
DyLight 488	492	517	Alternative
EGFP	488	507	Alternative
Emerald GFP	489	510	Alternative
FAM	492	518	Alte rnative
FITC	490	525	Alternative

Fluo-4





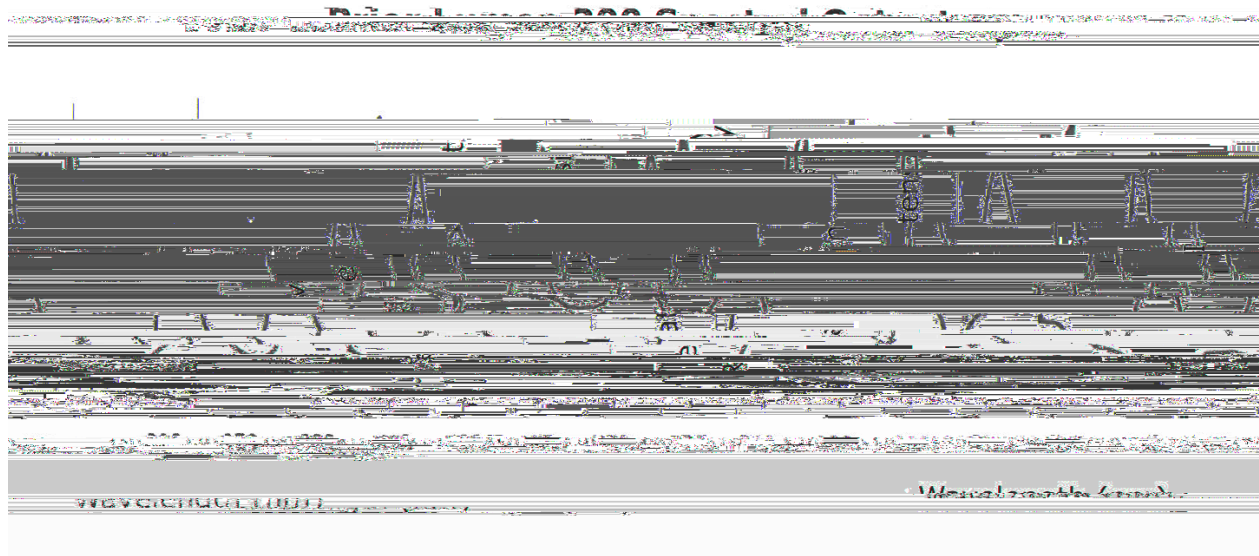
Position #5  
 49007 - ET - Cy7  
 Exciter ET710/75x  
 Beamsplitter T760LPXR  
 Emitter ET810/90m



-- ET710/75x -- | T7601pxr  
 -- ET810/90m

Fluorochrome	EX	EM	Use
Alexa Fluor 750TM	752		





## Hamamatsu O-1CA PZ Scint. Response

